

# DETECTION OF EXTENDED-SPECTRUM β-LACTAMASE PRODUCING ENTEROBACTERIACEAE **BY CHROMOGENIC ESBL SELECTIVE MEDIUM**

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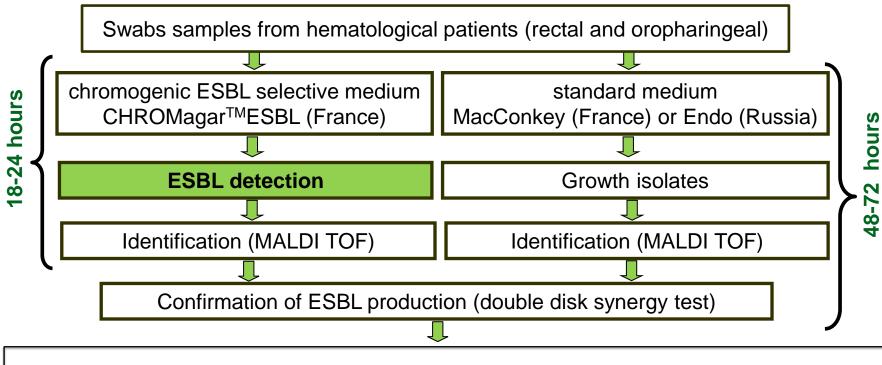
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# **OBJECTIVES**

The aim of the study was to evaluate the detection of extended-spectrum beta-lactamase producing Enterobacteriaceae (ESBL-E) by chromogenic ESBL selective medium and to compare results with double disk synergy test.

# **METHOD**

Prospective study was performed from April 2013 to December 2013. Study design was as follows:



#### AmpC detection

### **Additional tests**

Strains sensitive to Cefepime and resistant to Cefoxitin  $\rightarrow$  E-test (Cefotetan and Cefotetan with Cloxacillin)

Detection of TEM-1 and CTX-M by real-time PCR

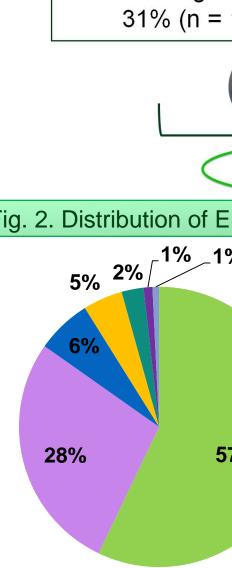
## RESULTS

We collected 1552 swabs and 1243 Enterobacteriaceae isolates were obtained. A total of 394 ESBL-E strains were recovered (Fig. 2), of those 123 (31%) only on chromogenic ESBL selective medium, 263 (67%) on standard medium and on CHROMagar<sup>™</sup>ESBL, 8 (2%) only on standard medium (p < 0,0001, Fig. 1). Characteristics of ESBL-E are summarized in Table 1. CTX-M group of ESBL were detected in the majority of ESBL-E.

Production of ESBL was confirmed for 94% (386/409) isolates obtained on chromogenic ESBL selective, other 6% (23/409) isolates were non-ESBL-E (Tab. 2). Sensitivity and specificity of CHROMagar<sup>™</sup>ESBL was 98% and 97%, respectively (Tab. 3).

# CONCLUSION

Chromogenic ESBL selective medium had high sensitivity (98%) and specificity (97%) for the detection of ESBL-E and might be used in routine laboratory practice. Detection of ESBL-E from the same rectal swabs by chromogenic ESBL selective medium was significantly higher than by standard medium (98% vs. 69%, p < 0,0001) and data were reported to clinical department in 18-24 hours.



RESULTS								
Fig. 1. ESBL-E detection								
ESBL-E N = 394								
	only on			CHROMagar™ESBL		only on standard		
	CHROMagar™ESBL			+ standard		medium		
	31% (n = 123)			67% (n=263)		2% (n = 8)		
98% (n = 386) p < 0,001 69% (n = 271)								
Fig. 2	Distribution	of ESBL-E	(N = 39)	Table 1. Characteristics of ESBL-E				
5% <sup>2% 1%</sup> 1%			■ E. coli (n=225)		ESBL-E	CTX-M, n (%)	TEM, n (%)	TEM+ CTX-M, n (%)
	69/		■ K. pneumoniae(n=109)		E. coli	157 (70)	107 (48)	64 (28)
	6%		<ul> <li>Enterobacter spp.(n=25)</li> <li>Citrobacter spp. (n=18)</li> </ul>		K. pneumoniae	51 (47)	36 (33)	10 (9)
					Enterobacter spp.	20 (80)	10 (40)	7 (28)
					Citrobacter spp.	14 (78)	8 (44)	7 (39)
2	28%	57%	■ K. oxytoca (n=10)		K. oxytoca P. mirabilis	7 (70)	3 (30)	2 (20)
			<ul> <li>P. mirabilis(n=4)</li> <li>Raoultella ornithinolytica (n=3)</li> </ul>		Raoultella	1 (25)	4 (100)	1 (25)
					ornithinolytica	2 (67)	1 (33)	1 (33)
					Total	252 (64)	169 (43)	92 (23)
	2. False positiv		of ESBL	Table 3. Sensitivity and specificity of CHROMagar <sup>™</sup> ESBL medium				
CHROMagar™ESBL (n=23)Strains (N)Speciesn (%)					Species of ESBL-E Sensitivity Specificity			
		Enterobacter spp.		11 (73)	E. coli	99%	99%	
omnC	(+) strains	Citrobacter spp.		2 (13)	K. pneumoniae	96%	100%	
(N=15		M. morganii		1 (7)	Other			
,		E. coli		1 (7)	Enterobacteriaceae	98%	95%	
	s sensitive to	E. coli		3 (37,5)	Total	98% 97%		97%
_		K. oxytoca		2 (25)	Notional Research Contex For Hemotoles			
cephalosporins (N=8)		P. vulgaris		2 (25)	National Research Center For Hematology, Novozykovski pr., 4, Moscow, 125167, Russia			
		Citrobacter spp.		1 (12,5)	E-mail: klyasova.g@blood.ru, amalofeeva@yandex.ru			



**EV0393**